

Article



Prevalence of Zinc Deficiency in Healthy 1–3-Year-Old Children from Three Western European Countries

Mirjam Vreugdenhil ^{1,*,†}, Marjolijn D. Akkermans ^{1,†}, Liandré F. van der Merwe ², Ruurd M. van Elburg ^{3,4}, Johannes B. van Goudoever ³ and Frank Brus ¹

- ¹ Department of Pediatrics, Juliana Children's Hospital, Haga Teaching Hospital, 2545 AA The Hague, The Netherlands; m.d.akkermans@hagaziekenhuis.nl (M.D.A.); fbrus1957@gmail.com (F.B.)
- ² Danone Nutricia Research, 3584 CT Utrecht, The Netherlands; liandre.vandermerwe@danone.com
- ³ Department of Pediatrics, Amsterdam UMC, Vrije Universiteit, University of Amsterdam Emma Children's Hospital, 1105 AZ Amsterdam, The Netherlands; rm.vanelburg@amsterdamumc.nl (R.M.v.E.); h.vangoudoever@amsterdamumc.nl (J.B.v.G.)
- ⁴ Nutriton4Health, 1214 LT Hilversum, The Netherlands
- Correspondence: m.vreugdenhil@hagaziekenhuis.nl
- + These authors contributed equally to this work.



Citation: Vreugdenhil, M.; Akkermans, M.D.; van der Merwe, L.F.; van Elburg, R.M.; van Goudoever, J.B.; Brus, F. Prevalence of Zinc Deficiency in Healthy 1–3-Year-Old Children from Three Western European Countries. *Nutrients* 2021, *13*, 3713. https:// doi.org/10.3390/nu13113713

Academic Editor: Elad Tako

Received: 8 September 2021 Accepted: 12 October 2021 Published: 22 October 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). **Abstract:** Zinc deficiency (ZnD) has adverse health consequences such as stunted growth. Since young children have an increased risk of developing ZnD, it is important to determine its prevalence and associated factors in this population. However, only a few studies have reported on ZnD prevalence in young children from Western high-income countries. This study evaluated ZnD prevalence and associated factors, including dietary Zn intake, in healthy 1–3-year-old children from Western European, high-income countries. ZnD was defined as serum Zn concentration <9.9 μ mol/L. A total of 278 children were included with a median age of 1.7 years (Q1–Q3: 1.2–2.3). The median Zn concentration was 11.0 μ mol/L (Q1–Q3: 9.0–12.2), and ZnD prevalence was 31.3%. No significant differences were observed in the socio-economic characteristics between children with and without ZnD. Dietary Zn intake was not associated with ZnD. ZnD is common in healthy 1–3-year-old children from Western European countries. However, the use of currently available cut-off values defining ZnD in young children has its limitations since these are largely based on reference values in older children. Moreover, these values were not evaluated in relation to health consequences, warranting further research.

Keywords: zinc; zinc deficiency; dietary zinc intake; children; Western Europe

1. Introduction

Zinc is an important trace element for human health, as it is involved in numerous and diverse biological processes throughout the body, including cell growth and differentiation, gene expression, and protein synthesis [1–3]. Subsequently, zinc deficiency (ZnD) has a wide range of possible adverse health consequences, including stunted growth, skin lesions and delayed wound healing, and impaired innate and adaptive immune functions [4–8].

ZnD is highly prevalent worldwide, resulting in substantial disease burden, especially in young children since they have an increased risk of developing ZnD due to increased requirements during periods of rapid growth [9,10]. In low- and middle-income countries, varying prevalence rates of ZnD have been reported in infants, young children, and preschool-age children, ranging from 5.1% in Sri Lanka to 82.6% in Cameroon [11]. Only a few studies report on the prevalence rates of ZnD in healthy young children from Western high-income countries, ranging from 0–60% [12–17]. However, most of these studies were conducted some time ago, while dietary habits in Western countries may have changed over the years. Moreover, in most of these studies, study populations were relatively small, and associated factors were not extensively assessed. Because of the possible adverse health consequences of ZnD in young children, it is important to determine ZnD prevalence and associated factors in this population to implement and guide preventive strategies. Therefore, this study aimed to assess the prevalence of ZnD in a population of healthy 1–3-year-old children living in Western Europe and evaluate the possible associated factors, including dietary Zn intake.

2. Materials and Methods

2.1. Study Design

Data were used from a previously conducted randomized, double-blind controlled trial (NTR3609) investigating the effect of a micronutrient-fortified growing-up milk given for 20 weeks on the iron and vitamin D status of healthy 1–3-year-old children in comparison to non-fortified cow's milk [18]. This study was conducted from October 2012 to September 2014 in three Western European countries, namely Germany (nine private pediatric clinics), the Netherlands (one secondary and two tertiary hospitals), and the United Kingdom (UK) (one secondary and one tertiary hospital). The study protocol was approved by the medical ethical review boards of all participating sites and has been previously described in detail [18]. In addition to iron and vitamin D status, other micronutrients, including zinc (Zn), were also analyzed. Blood samples collected during the baseline study visit were used for the current study.

2.2. Study Population and Procedure

Children between 1 and 3 years of age with a stable health status (i.e., no known chronic or recent acute diseases, no known infection during the last week, or infection needing medical assistance or treatment during the last two weeks) and expected to remain stable were eligible for participation. The recruitment of participants differed between countries. In the Netherlands and the UK, parent(s) or legal representative(s) of eligible children were informed about the study during a preoperative visit before an elective non-emergency surgical procedure (e.g., urologic surgeries, inguinal or umbilical hernia operations, or ear-nose-throat procedures). If written informed consent was obtained from the parent(s)/legal representative(s), the baseline study visit coincided with the elective non-emergency surgical procedure. The baseline blood sample was then combined with intravenous injection necessary for administering general anesthesia. In Germany, participants were recruited during a regular visit to a pediatric clinic. During the baseline study visit, the child's body weight and height/length were measured. Body weight was measured while wearing only underwear, using a calibrated weighing scale. Height was measured while standing and without wearing shoes, using a calibrated stadiometer. Length was measured in children for whom it was not possible to take a height measurement while standing, using a length board. Weight-for-age-z-scores and height/lengthfor-age-z-scores were calculated using World Health Organization (WHO) Child Growth Standards. Stunting was defined as the height/length-for-age >2 standard deviations (SD) below the median of WHO Child Growth Standards [19]. Moreover, parent(s)/legal representative(s) were asked questions during the baseline visit about their child's demographicand socio-economic characteristics, daycare center attendance, medical history, and dietary intake [18]. Dietary intake was estimated using a food-frequency questionnaire adapted from other dietary questionnaires [20,21], as previously described [18]. This questionnaire reflects dietary intake during the month before the baseline visit. For this study, we mainly focused on the food groups known to affect zinc status (e.g., meat, vegetables, fish, vegetables, grains and grain-based products, and eggs). Zn intake (mg/day) (per reported food group) was determined by using the Dutch Food Composition Database [22].

2.3. Laboratory Analysis and Definitions

Venous blood samples were collected from participants throughout various times of the day by trained personnel using trace-element free heparinized tubes, following local laboratory protocol and procedures recommended by the International Zinc Nutrition Consultative Group (IZiNCG) [19]. After clotting and centrifugation, serum was distributed and aliquoted in polypropylene tubes and stored between -20 °C and -80 °C at the study site. From the study sites, samples were shipped (within two months) on dry ice to the Nutricia Research Analytical Science Laboratory in Utrecht, the Netherlands, where samples were stored at -80 °C. From the Nutricia Research Analytical Science Laboratory, samples were shipped on dry ice to the Reinier Haga Medical Diagnostic Center in Delft, the Netherlands, where samples were stored at -80 °C until analysis. Serum Zn concentrations were determined using flame atomic absorption spectrophotometry (AA-7000, Shimadzu) and hsCRP concentrations using a clinical chemistry analyzer (Abbott Architect c16000) with a turbidimetry method. ZnD was defined as a serum Zn concentration <9.9 μ mol/L [19]. Since Zn concentration may be reduced in the case of infection or inflammation [23–25], participants with a high-sensitivity C-reactive protein (hsCRP) concentration >5 mg/L [26] were excluded from analyses. Additionally, children with missing Zn and/or hsCRP measurements at baseline were also excluded from analyses.

2.4. Statistical Analysis

SPSS version 24.0 (SPSS Inc., Chicago, IL, USA) was used for all analyses. Distribution of data was assessed using histograms and Shapiro–Wilk tests. Data are presented as the mean with SD for normally distributed variables or median with first and third quartiles (Q1–Q3) for non-normally distributed variables. Categorical variables are presented as numbers with percentages. Comparisons between groups were made using the independent T-test for normally distributed variables and the Mann- Whitney U non-parametric test for non-normally distributed variables. For categorical variables, comparisons between groups were made using the Chi-squared or Fisher's exact test. Statistical significance was defined as p < 0.05. In order to further explore factors associated with ZnD, a binary logistic regression analysis was performed using the variables previously identified with a p-value < 0.1 as covariates, including age and sex independent of p-values, and ZnD as dependent variable.

3. Results

3.1. Study Population

In total, 278 of the 325 (85.5%) healthy 1–3-year-old children of the original nutritional intervention study were included for our Zn analyses: 237 (85.3%) in Germany, 37 (13.3%) in the Netherlands, and 4 (1.4%) in the UK. Children excluded from analyses had missing Zn and/or hsCRP measurements or an hsCRP concentration >5 mg/L at baseline. There were no statistically significant differences regarding socio-economic characteristics and dietary intake found between included and excluded children (data not shown). The median age of the study participants was 1.7 years (1.2–2.3). Of the study population, 153 children (55.0%) were male, and 267 (96.0%) were Caucasian. Stunting was present in three children (1.1%). More characteristics of the study population are shown in Table 1.

	$\begin{array}{c} \mathbf{A11}\\ \mathbf{N} = 278 \end{array}$	ZnD N = 87	No ZnD N = 191	р
Characteristics				
Age (years)	1.7 (1.2–2.3)	1.6 (1.1–2.3)	1.7 (1.2–2.3)	0.301
Sex (male)	153 (55.0%)	48 (55.2%)	105 (55.0%)	0.975
Ethnicity (Caucasian)	267 (96.0%)	85 (97.7%)	182 (95.3%)	0.511
From Germany	237 (85.3%)	86 (98.9%)	151 (79.1%)	< 0.001 *
Parental educational level At least one parent with university education Neither with university education	(N = 231, M = 47) 53 (19.1%) 178 (64.0%)	(N = 74, M = 13) 16 (18.4%) 58 (66.7%)	(N = 157, M = 34) 38 (19.4%) 120 (62.8%)	0.743
Parental professional status At least one parent working Neither working	(N = 219, M = 59) 210 (75.5%) 9 (3.2%)	(N = 69, M = 18) 67 (77.0%) 2 (2.3%)	(N = 150, M = 41) 143 (74.9%) 7 (3.7%)	0.723

Table 1. Characteristics and dietary intake of the study population (1-3 years of age) and relation to Zn status.

	$\begin{array}{c} A11\\ N=278 \end{array}$	ZnD N = 87	No ZnD N = 191	р
Characteristics				
Daycare attendance	117 (42.1%) (N = 276, M = 2)	36 (41.4%)	81 (42.4%) (N = 189, M = 2)	0.817
Weight-for-age-z-score	0.22 (±0.94) (N = 276, M = 2)	0.20 (±0.99)	0.24 (±0.92) (N = 189, M = 2)	0.727
Height/length-for-age-z-score	-0.03 (±0.99) (N = 271, M = 7)	0.04 (±1.04)	$-0.07 (\pm 0.97)$ (N = 184, M = 7)	0.417
Stunting	3 (1.1%) (N = 271, M = 7)	1 (1.1%)	2 (1.0%) (N = 184, M = 7)	NA
Dietary intake				
Ever breastfed [§]	177 (63.7%) (N = 267, M = 11)	64 (73.6%)	113 (59.2%) (N = 180, M = 11)	0.081
Duration of breastfeeding § 0-<6 months ≥ 6 months	102 (57.6%) 75 (42.4%)	34 (53.1%) 30 (46.9%)	68 (60.2%) 45 (39.8%)	0.362
Ever formula fed [§]	251 (90.3%) (N = 267, M = 11)	79 (90.8%)	172 (90.1%) (N = 180, M = 11)	0.125
Age of introduction of solid foods (months) 0–6 months >6 months	(N = 273, M = 5) 224 (80.6%) 49 (17.6%)	(N = 86, M = 1) 74 (85.1%) 12 (13.8%)	(N = 187, M = 4) 150 (78.5%) 37 (19.4%)	0.243
Main type of milk intake [†] ° Use of primarily cow's milk Use of primarily formula	(N = 268, M = 10) 119 (42.8%) 147 (52.9%)	40 (46.0%) 47 (54.0%)	(N = 181, M = 10) 79 (41.4%) 100 (52.4%)	0.719 0.850
Use of dietary supplements (unknown zinc content)	86 (30.9%)	27 (31.0%)	59 (30.9%)	0.798
Total amount of milk per day ° (mL/day) Zn intake from milk in general (mg/day)	420 (400–600) 2.78 (1.95–3.57) (N = 268, M = 10)	420 (300–600) 2.76 (2.05–3.34)	420 (400–600) 2.79 (1.84–3.68) (N = 181, M = 10)	0.376 0.481
Meat (g/day) Zn intake from meat (mg/day)	19.3 (11.7–29.7) 0.63 (0.38–0.96) (N = 268, M = 10)	19.3 (11.9–29.7) 0.63 (0.38–0.96)	18.8 (11.0–29.7) 0.61 (0.36–0.96) (N = 181, M = 10)	0.611 0.611
Fish (g/day) Zn intake from fish (mg/day)	2.3 (0.9–4.5) 0.02 (0.01–0.03) (N = 268, M = 10)	2.4 (1.2–4.7) 0.02 (0.01–0.04)	2.0 (0.8–4.4) 0.02 (0.01–0.03) (N = 181, M = 10)	$0.458 \\ 0.458$
Vegetables (g/day) Zn intake from vegetables (mg/day)	61.0 (35.3–112.4) 0.57 (0.33–1.05) (N = 268, M = 10)	63.0 (35.9–132.7) 0.59 (0.34–1.25)	59.3 (34.6–108.1) 0.56 (0.32–1.01) (N = 181, M = 10)	0.369 0.369
Dried fruits, seeds and nuts (g/day) Zn intake from dried fruits, seeds and nuts	0.6 (0.0–5.8) 0.01 (0.00–0.12)	0.0 (0.0–3.6) 0.00 (0.00–0.07)	0.8 (0.0–6.2) 0.02 (0.00–0.12)	0.068 0.068
(mg/day)	(N = 268, M = 10)		(N = 181, M = 10)	
Bread (g/day) Zn intake from bread (mg/day)	33.7 (32.4–64.9) 0.43 (0.41–0.83) (N = 268, M = 10)	33.7 (32.4–64.9) 0.43 (0.41–0.83)	46.2 (23.8–64.9) 0.59 (0.30–0.83) (N = 181, M = 10)	0.415 0.415
Sandwich spread (g/day) Zn intake from sandwich spread (mg/day)	1.1 (0.0–4.5) 0.01 (0.00–0.03) (N = 267, M = 11)	0.9 (0.0–3.5) 0.01 (0.00–0.03)	1.1 (0.1–4.8) 0.01 (0.00–0.04) (N = 180, M = 11)	0.436 0.436
Breakfast cereals (g/day) Zn intake from breakfast cereals (mg/day)	3.0 (0.0–24.1) 0.06 (0.00–0.47) (N = 268, M = 10)	5.9 (0.0–32.5) 0.12 (0.00–0.64)	1.9 (0.0–23.0) 0.04 (0.00–0.45) (N = 181, M = 10)	0.230 0.230
Eggs (g/day) Zn intake from eggs (mg/day)	6.7 (1.6–6.7) 0.10 (0.02–0.10) (N = 268, M = 10)	6.7 (1.6–6.7) 0.10 (0.02–0.10)	3.9 (1.6–6.7) 0.06 (0.02–0.10) (N = 181, M = 10)	0.591 0.591
Zn intake from solid foods (mg/day)	2.52 (1.86–3.73) (N = 268, M = 10)	2.76 (2.20–3.73)	2.33 (1.77–3.75) (N = 181, M = 10)	0.066
Total dietary Zn intake (mg/day) (from milk and solid foods)	5.71 (4.43–6.85) (N = 268, M = 10)	5.59 (4.57-6.74)	5.83 (4.33–6.93) (N = 181, M = 10)	0.978

Data are expressed as medians (Q1–Q3) or numbers (percentages). In the case of normal distribution, means (standard deviations) are reported. [§] Exclusively or partially. [†] During the previous month; [°] N = 2 other main types of milk/dairy (i.e., human milk (N = 1), yogurt (N = 1)). *p* denotes *p*-value regarding comparisons between "ZnD" and "no ZnD". * Statistically significant with *p* < 0.05. Abbreviations: Zn, zinc; ZnD, zinc deficiency; N, number; M, missing; NA, not applicable.

Table 1. Cont.

3.2. Serum Zn Concentrations, ZnD Prevalence and Dietary Zn Intake

In the total study population, the median Zn concentration was 11.0 μ mol/L (9.0–12.2). The median Zn concentrations in children with and without ZnD were 9.0 μ mol/L (8.0–9.0) and 12.0 μ mol/L (10.7–13.0), respectively. ZnD was present in 87 (31.3%) children. The median total dietary Zn intake (from milk and solid foods) was 5.71 mg/day (4.43–6.85), with a median Zn intake from milk of 2.78 mg/day (1.95–3.57) and from solid foods of 2.52 mg/day (1.86–3.73). More dietary intake details of the study population are shown in Table 1.

3.3. Factors Associated with ZnD

No significant differences in sex distribution, ethnicity, parental educational or professional status, or daycare attendance were found between children with and without ZnD. Children with ZnD were significantly more often 'from Germany' (98.9%) compared to children without ZnD (79.1%, p < 0.001). Furthermore, no significant differences in the consumption of solid foods (gr/day) or Zn intake (mg/day) per reported food group in children with and without ZnD were found. Finally, no significant differences in Zn intake from milk (i.e., formula, cow's milk, or otherwise) (p = 0.481), solid foods (p = 0.066), or total dietary Zn intake (from milk and solid foods) (p = 0.978) between children with and without ZnD were observed (Table 1).

A binary logistic regression analysis was performed to further explore factors associated with ZnD, with 'ZnD' as dependent variable and 'age', 'sex', 'ever breastfed', 'from Germany', and 'dietary Zn intake' as covariates, which showed that being 'from Germany' was significantly associated with ZnD (OR = 16.6; 95% CI 92.2–124.2; p = 0.006). No other significant associations with ZnD were found (data not shown). Although 'Zn intake from dried fruits, seeds and nuts' had a p-value < 0.1, we chose not to include this variable in our multivariate model since this variable was already included in 'dietary Zn intake'. A separate binary logistic analysis with 'Zn intake from dried fruits, seeds and nuts' instead of 'dietary Zn intake', showed no significant association with ZnD (data not shown).

A subgroup analysis was performed in 1–2-year-old and 2–3-year-old children: ZnD was present in 54 (30.2%) and 33 (33.3%) of 1–2-year and 2–3-year-old children, respectively, and this difference in ZnD prevalence was not significant (p = 0.586).

4. Discussion

This study showed that ZnD, with a prevalence rate of 31.3%, is common in healthy 1–3-year-old children living in three Western European countries. In addition to being from Germany, no other evaluated factors, including total dietary Zn intake, were associated with ZnD.

4.1. ZnD Prevalence

With an overall prevalence of 31%, we showed that ZnD is not uncommon in healthy 1–3-year-old children living in three Western European high-income countries. In lowand middle-income countries, prevalence rates of ZnD between 5% and 83% have been reported in young children [11]. Populations in low- and middle-income countries are at an increased risk of inadequate Zn intake and subsequently ZnD, which can be partially attributed to limited access to foods that are rich in zinc, such as animal products, in combination with a mainly plant-based diet, which contains phytates that inhibit intestinal Zn absorption [27]. Studies conducted in young children in Western high-income countries also report varying prevalence rates of ZnD, ranging between 0% and 60% in 1-year-old children [12,14,15,17]. ZnD was found in 21% of healthy children below three years of age in France [13] and in 38% of healthy toddlers between 12 and 20 months of age in New Zealand [16]. However, the comparison of prevalence rates of ZnD between studies remains challenging due to relatively small sample sizes and differences in study populations.

4.2. Factors Associated with ZnD

In our study population, we found no association between ZnD and the evaluated sociodemographic factors, besides being from Germany. Children with ZnD were significantly more often 'from Germany' (98.9%) compared to children without ZnD (79.1%). We cannot rule out that this association is possibly due to differences in setting and recruitment procedures (e.g., fasting state) between children recruited from Germany and children recruited from the Netherlands and the UK. However, we believe that since the majority of participants were from Germany (85%), it is likely by chance that children with ZnD are more likely to come from Germany than from the Netherlands or the UK. Moreover, no association between ZnD and dietary Zn intake was found. Three meta-analyses also concluded that serum Zn concentrations are unrelated to Zn intake [28–30]. Furthermore, Hennigar et al. [31] reported no differences in serum Zn concentrations between those with lower or higher Zn intake than the recommended dietary allowance for Zn. With a median total dietary Zn intake of 5.6 mg/day, our study population meets the recommended Zn intakes in each participating country [32–34]. The lack of association between dietary Zn intake and Zn concentration might be explained by factors that influence intestinal Zn absorption [35–37]. As mentioned before, phytates inhibit Zn absorption [27], whereas an increased amount of dietary protein, for example, increases Zn absorption [38,39]. The type of dietary protein also affects Zn absorption, for example, animal-derived proteins increase Zn absorption compared to plant-derived proteins [40,41]. In short, the composition of the diet might impact intestinal Zn absorption, despite sufficient Zn intake, and this might (partially) explain why we found no association between ZnD and dietary Zn intake.

4.3. Strengths and Limitations

The strengths of this study are the relatively large study population of healthy 1–3-year-old, mainly Caucasian, children from three Western European countries, and the effect of inflammation and/or infection on Zn concentrations that was taken into account by excluding children with CRP > 5 mg/L. It is known that Zn concentrations can be affected by factors such as inflammation, fasting state, and/or diurnal rhythm [42]. A limitation of this study is that we did not have information concerning the fasting state during blood draw, the time of blood draw, and meal consumption prior to the blood draw, which might have influenced the reported Zn concentrations [31]. The cut-off value of 9.9 µmol/L $(65 \,\mu g/dL)$ used to define ZnD, as suggested by the IZiNCG, accounts for blood samples collected in the morning with a non-fasting state. For blood samples collected in the afternoon, the IZiNCG suggests a cut-off value of 8.7 μ mol/L (57 μ g/dL) [19]. Since we do not know the exact time the blood samples were collected, we chose the highest cut-off value for a non-fasting state (i.e., 9.9 µmol/L) to define ZnD, which may have resulted in an overestimation of the prevalence of ZnD in our study population. However, when using the cut-off value of 57 μ g/dL (8.7 μ mol/L) for afternoon samples, we found the prevalence of ZnD to be 11.5%, which is still relatively high.

4.4. Defining ZnD

Finally, we want to address some concerns regarding the current definition of ZnD. At present, Zn concentration in either plasma or serum is the most extensively investigated and widely used biomarker to assess zinc status. Currently, the cut-off values for determining ZnD are largely based on reference values derived from large cohorts of presumably healthy people, such as the second National Health and Nutrition Examination Survey (NHANES II) during the period 1976–1980 [43]. Based on the results from the NHANES II survey, cut-off values for ZnD are suggested by the IZiNCG, i.e., <65 μ g/dL (9.9 μ mol/L) in the morning, and <57 μ g/dL (8.7 μ mol/L) in the afternoon for children below ten years of age [19], and are widely used in most studies evaluating zinc status in humans. However, children under the age of three were not represented in this survey. It is, therefore, possible that these cut-off values do not apply for 1–3-year-old children. Two studies evaluated serum Zn concentrations in healthy 1–3-year-old children living in Australia

and Belgium [44,45]. First, reference intervals (2.5th and 97.5th percentiles) for serum Zn concentrations reported by Karr et al. were 9–19 μ mol/L and 8–19 μ mol/L in 1–2-year-old and 2–3-year-old healthy Australian children [44]. Second, van Biervliet et al. found mean serum Zn concentrations of 12.7 μ mol/L (SD 2.8 μ mol/L) in 1–2-year-old children and 12.7 μ mol/L (SD 3.1 μ mol/L) in 2–3-year-old healthy Belgian children [45,46]. These values do not differ that much from those resulting from the NHANES II survey in 3–4-year-old children (i.e., 12.2 μ mol/L (SD 2.0 μ mol/L)) [46]. However, both studies did not clearly take into account the presence of infection or the time of blood sampling which both can influence Zn concentration [44,45].

However, Zn concentration might not accurately reflect zinc status. Zn is present in all tissues, of which skeletal muscle and bone have the highest zinc content (approximately 83% of total body zinc), whereas plasma accounts for less than 0.2% of total body zinc. It is currently unknown whether Zn concentrations below a certain predefined cut-off value (e.g., based on cohort-derived reference values) fail to meet bodily requirements and cause negative health consequences. In adults, a lower cut-off value of 50 µg/dL (i.e., 7.6 µmol/L) has been proposed for clinical ZnD associated with symptoms, including diarrhea and alopecia [47]. To our knowledge, no such studies have been conducted in (young) children (to date). In our study, stunting (i.e., a functional health outcome of ZnD) was only prevalent in three children (1.1%), and due to this small number, no statistical analysis was performed regarding a possible association with ZnD. Furthermore, other biomarkers for assessing Zn status have been evaluated. For example, urinary zinc excretion, hair zinc concentration [48], or zinc-binding proteins, such as metallothionein [42], might be promising biomarkers for assessing human zinc status. However, due to a limited number of studies and subgroups within these studies, results are difficult to apply in clinical practice. More research is needed to obtain more insight into the use of these biomarkers, preferably in relation to clinical symptoms, and to identify more direct biomarkers for assessing Zn status.

5. Conclusions

ZnD is highly prevalent in healthy 1–3-year-old children in Western Europe. No significant differences were observed in the socio-economic characteristics between children with and without ZnD. Dietary Zn intake was not associated with ZnD. The latter might partially be explained by various dietary factors influencing intestinal Zn absorption. Further studies are needed regarding the influence of diet composition on Zn absorption in young children. Moreover, future research is required to assess reference values for Zn concentrations in relation to clinical symptoms and health outcomes in young children and gain more insight into the use of certain other proposed and promising biomarkers, such as metallothionein.

Author Contributions: M.V., M.D.A., L.F.v.d.M., R.M.v.E., J.B.v.G., and F.B. designed the research; M.V. and M.D.A. implemented and conducted the research; M.V. and M.D.A. performed the statistical analyses; M.V., M.D.A., and F.B. prepared and wrote the original draft; M.D.A., L.F.v.d.M., R.M.v.E., J.B.v.G., and F.B. reviewed the manuscript. All authors critically revised the manuscript; All authors have read and agreed to the published version of the manuscript.

Funding: The clinical trial leading to these results was financially supported by Danone Nutricia Research. Statistical analyses and interpretation of the data were performed independently. The current research received no external funding.

Institutional Review Board Statement: This study was conducted according to the guidelines of the Declaration of Helsinki, and approved by the Institutional Review Board (or Ethics Committee) of the Medical Ethics Committee of South-West Holland (protocol code: 12-058—NL41045.098.12 and date of approval: 20 September 2012); National Research Ethics Service London (protocol code: Tod.1.C.L. and date of approval: 8 November 2012); Ethics Committee Niedersachsen (protocol code: Tod.1.C.L. and date of approval: 29 October 2012); Ethics Committee Thüringen (protocol code: Tod.1.C.L. and date of approval: 22 April 2013); Ethics Committee Thüringen (protocol code: Tod.1.C.L. and date of approval: 23 November 2012); Ethics Committee Hessen (protocol code: Tod.1.C.L. and date of approval: 23 November 2012); Ethics Committee Hessen (protocol code: Tod.1.C.L. and date of approval: 20 September 2012); Ethics Committee Hessen (protocol code: Tod.1.C.L. and date of approval: 23 November 2012); Ethics Committee Hessen (protocol code: Tod.1.C.L. and date of approval: 20 September 2012); Ethics Committee Hessen (protocol code: Tod.1.C.L. and date of approval: 23 November 2012); Ethics Committee Hessen (protocol code: Tod.1.C.L. and date of approval: 24 Protocol 2012); Ethics Committee Hessen (protocol code: Tod.1.C.L. and date of approval: 24 Protocol 2012); Ethics Committee Hessen (protocol code: Tod.1.C.L. and date of approval: 25 November 2012); Ethics Committee Hessen (protocol code: Tod.1.C.L. and date of approval: 26 Protocol 2012); Ethics Committee Hessen (protocol code: Tod.1.C.L. and date of approval: 26 Protocol 2012); Ethics Committee Hessen (protocol code: Tod.1.C.L. and date of approval: 27 Protocol 2012); Ethics Committee Hessen (protocol code: Tod.1.C.L. and date of approval: 28 Protocol 2012); Ethics Committee Hessen (protocol code: Tod.1.C.L. and date of approval: 29 Protocol 2012); Ethics Committee Hessen (protocol code: Tod.1.C.L. and date of approval: 29 Protocol 2012); Ethics Protocol 2012); Ethics Protoc

approval: 7 December 2012); Ethics Committee Nordrhein Westfalen (protocol code: Tod.1.C.L. and date of approval: 28 December 2012).

Informed Consent Statement: Written informed consent was obtained from parent(s)/legal representative(s) of all participants.

Data Availability Statement: The data presented in this study are available on request from the corresponding author. The data are not publicly available since the participants of this study did not agree for their data to be shared publicly.

Acknowledgments: We thank the children and parents who participated in the initial clinical trial. Additionally, we thank all the pediatricians involved for their contributions to the initial clinical trial, and therefore to this study.

Conflicts of Interest: L.F.v.d.M. is an employee of Danone Nutricia Research. R.M.v.E. was an employee of Danone Nutricia Research when the initial clinical trial was conducted. They had no role in the execution of the study or the statistical analyses of the results. J.B.v.G. is member of the National Health Council and founder and director of the National Human Milk Bank. He received financial support for studies from and through the EU (FP7 & H2020, Danone). None of the other authors reported a conflict of interest related to the study.

References

- 1. Vallee, B.L.; Falchuk, K.H. The biochemical basis of zinc physiology. *Physiol. Rev.* 1993, 73, 79–118. [CrossRef]
- 2. Cousins, R.J. Metal elements and gene expression. Annu. Rev. Nutr. 1994, 14, 449–469. [CrossRef]
- 3. MacDonald, R.S. The role of zinc in growth and cell proliferation. J. Nutr. 2000, 130, 1500s–1508s. [CrossRef]
- 4. Imdad, A.; Bhutta, Z.A. Effect of preventive zinc supplementation on linear growth in children under 5 years of age in developing countries: A meta-analysis of studies for input to the lives saved tool. *BMC Public Health* **2011**, *11* (Suppl. 3), S22. [CrossRef]
- 5. Prasad, A.S. Discovery of human zinc deficiency: Its impact on human health and disease. Adv. Nutr. 2013, 4, 176–190. [CrossRef]
- 6. Liu, E.; Pimpin, L.; Shulkin, M.; Kranz, S.; Duggan, C.P.; Mozaffarian, D.; Fawzi, W.W. Effect of zinc supplementation on growth outcomes in children under 5 years of age. *Nutrients* **2018**, *10*, 377. [CrossRef]
- 7. Ogawa, Y.; Kinoshita, M.; Shimada, S.; Kawamura, T. Zinc and skin disorders. Nutrients 2018, 10, 199. [CrossRef] [PubMed]
- 8. Gammoh, N.Z.; Rink, L. Zinc in Infection and Inflammation. *Nutrients* 2017, 9, 624. [CrossRef]
- 9. Gibson, R.S. Zinc: The missing link in combating micronutrient malnutrition in developing countries. *Proc. Nutr. Soc.* 2006, 65, 51–60. [CrossRef] [PubMed]
- Mayo-Wilson, E.; Junior, J.A.; Imdad, A.; Dean, S.; Chan, X.H.; Chan, E.S.; Jaswal, A.; Bhutta, Z.A. Zinc supplementation for preventing mortality, morbidity, and growth failure in children aged 6 months to 12 years of age. *Cochrane Database Syst. Rev.* 2014, Cd009384. [CrossRef] [PubMed]
- Gupta, S.; Brazier, A.K.M.; Lowe, N.M. Zinc deficiency in low- and middle-income countries: Prevalence and approaches for mitigation. J. Hum. Nutr. Diet. Off. J. Br. Diet. Assoc. 2020, 33, 624–643. [CrossRef] [PubMed]
- 12. Persson, L.A.; Lundström, M.; Lönnerdal, B.; Hernell, O. Are weaning foods causing impaired iron and zinc status in 1-year-old Swedish infants? A cohort study. *Acta Paediatr. (Oslo, Norway: 1992)* **1998**, *87*, 618–622. [CrossRef]
- 13. Bouglé, D.; Laroche, D.; Bureau, F. Zinc and iron status and growth in healthy infants. *Eur. J. Clin. Nutr.* 2000, *54*, 764–767. [CrossRef] [PubMed]
- 14. Kattelmann, K.K.; Ho, M.; Specker, B.L. Effect of timing of introduction of complementary foods on iron and zinc status of formula fed infants at 12, 24, and 36 months of age. *J. Am. Diet. Assoc.* **2001**, *101*, 443–447. [CrossRef]
- 15. Taylor, A.; Redworth, E.W.; Morgan, J.B. Influence of diet on iron, copper, and zinc status in children under 24 months of age. *Biol. Trace. Elem. Res.* **2004**, *97*, 197–214. [CrossRef]
- Morgan, E.J.; Heath, A.L.; Szymlek-Gay, E.A.; Gibson, R.S.; Gray, A.R.; Bailey, K.B.; Ferguson, E.L. Red meat and a fortified manufactured toddler milk drink increase dietary zinc intakes without affecting zinc status of New Zealand toddlers. *J. Nutr.* 2010, 140, 2221–2226. [CrossRef]
- 17. Daniels, L.; Williams, S.M.; Gibson, R.S.; Taylor, R.W.; Samman, S.; Heath, A.M. Modifiable "predictors" of zinc status in toddlers. *Nutrients* **2018**, *10*, 306. [CrossRef] [PubMed]
- Akkermans, M.D.; Eussen, S.R.; van der Horst-Graat, J.M.; van Elburg, R.M.; van Goudoever, J.B.; Brus, F. A micronutrientfortified young-child formula improves the iron and vitamin D status of healthy young European children: A randomized, double-blind controlled trial. *Am. J. Clin. Nutr.* 2017, 105, 391–399. [CrossRef]
- 19. International Zinc Nutrition Consultative Group. *IZiNCG Technical Brief* 2007: Assessing Population Zinc Status with Serum Zinc Concentration; International Zinc Nutrition Consultative Group (IZiNCG): Davis, CA, USA, 2007.
- Geleijnse, J.M.; Giltay, E.J.; Schouten, E.G.; de Goede, J.; Oude Griep, L.M.; Teitsma-Jansen, A.M.; Katan, M.B.; Kromhout, D. Effect of low doses of n-3 fatty acids on cardiovascular diseases in 4837 post-myocardial infarction patients: Design and baseline characteristics of the Alpha Omega Trial. Am. Heart J. 2010, 159, 539–546.e532. [CrossRef]

- Uijterschout, L.; Vloemans, J.; Vos, R.; Teunisse, P.P.; Hudig, C.; Bubbers, S.; Verbruggen, S.; Veldhorst, M.; de Leeuw, T.; van Goudoever, J.B.; et al. Prevalence and risk factors of iron deficiency in healthy young children in the southwestern Netherlands. *J. Pediatric Gastroenterol. Nutr.* 2014, *58*, 193–198. [CrossRef]
- Rijksinstituut voor Volksgezondheid en Milieu (RIVM) (Dutch Institute for Public Health and Environment). Nederlands Voedingsstoffenbestand (NEVO) (Dutch Food Composition Database). Available online: https://www.rivm.nl/nederlandsvoedingsstoffenbestand (accessed on 2 February 2021).
- 23. Galloway, P.; McMillan, D.C.; Sattar, N. Effect of the inflammatory response on trace element and vitamin status. *Ann. Clin. Biochem.* **2000**, *37 Pt 3*, 289–297. [CrossRef]
- 24. Wieringa, F.T.; Dijkhuizen, M.A.; West, C.E.; Muhilal, C.A.N.-C. Estimation of the effect of the acute phase response on indicators of micronutrient status in Indonesian infants. *J. Nutr.* **2002**, *132*, 3061–3066. [CrossRef] [PubMed]
- 25. Mburu, A.S.; Thurnham, D.I.; Mwaniki, D.L.; Muniu, E.M.; Alumasa, F.M. The influence of inflammation on plasma zinc concentration in apparently healthy, HIV+ Kenyan adults and zinc responses after a multi-micronutrient supplement. *Eur. J. Clin. Nutr.* **2010**, *64*, 510–517. [CrossRef] [PubMed]
- 26. International Zinc Nutrition Consultative Group. *IZiNCG Technical Brief* 2020: *Adjusting Plasma or Serum Zinc Concentrations for Inflammation;* International Zinc Nutrition Consultative Group (IZiNCG): Davis, CA, USA, 2020.
- 27. Lönnerdal, B. Dietary factors influencing zinc absorption. J. Nutr. 2000, 130, 1378s–1383s. [CrossRef] [PubMed]
- Lowe, N.M.; Medina, M.W.; Stammers, A.L.; Patel, S.; Souverein, O.W.; Dullemeijer, C.; Serra-Majem, L.; Nissensohn, M.; Hall Moran, V. The relationship between zinc intake and serum/plasma zinc concentration in adults: A systematic review and dose-response meta-analysis by the EURRECA Network. *Br. J. Nutr.* 2012, *108*, 1962–1971. [CrossRef] [PubMed]
- Moran, V.H.; Stammers, A.L.; Medina, M.W.; Patel, S.; Dykes, F.; Souverein, O.W.; Dullemeijer, C.; Pérez-Rodrigo, C.; Serra-Majem, L.; Nissensohn, M.; et al. The relationship between zinc intake and serum/plasma zinc concentration in children: A systematic review and dose-response meta-analysis. *Nutrients* 2012, *4*, 841–858. [CrossRef] [PubMed]
- 30. King, J.C. Yet again, serum zinc concentrations are unrelated to zinc intakes. J. Nutr. 2018, 148, 1399–1401. [CrossRef]
- 31. Hennigar, S.R.; Lieberman, H.R.; Fulgoni, V.L., 3rd; McClung, J.P. Serum zinc concentrations in the us population are related to sex, age, and time of blood draw but not dietary or supplemental zinc. *J. Nutr.* **2018**, *148*, 1341–1351. [CrossRef]
- 32. Haase, H.; Ellinger, S.; Linseisen, J.; Neuhäuser-Berthold, M.; Richter, M. Revised D-A-CH-reference values for the intake of zinc. *J. Trace Elem. Med. Biol. Organ Soc. Miner. Trace Elem.* (*GMS*) **2020**, *61*, 126536. [CrossRef]
- Foundation, B.N. Nutrition Requirements. Available online: https://www.nutrition.org.uk/attachments/article/907/Nutrition% 20Requirements_Revised%20August%202019.pdf (accessed on 9 April 2021).
- 34. Voedingscentrum. Zink. Available online: https://www.voedingscentrum.nl/encyclopedie/zink.aspx (accessed on 9 April 2021).
- 35. Maares, M.; Haase, H. A guide to human zinc absorption: General overview and recent advances of in vitro intestinal models. *Nutrients* **2020**, *12*, 762. [CrossRef]
- 36. King, J.C.; Shames, D.M.; Woodhouse, L.R. Zinc homeostasis in humans. J. Nutr. 2000, 130, 1360s–1366s. [CrossRef] [PubMed]
- 37. Hambidge, M.; Krebs, N.F. Interrelationships of key variables of human zinc homeostasis: Relevance to dietary zinc requirements. *Annu. Rev. Nutr.* **2001**, *21*, 429–452. [CrossRef] [PubMed]
- Sandström, B.; Arvidsson, B.; Cederblad, A.; Björn-Rasmussen, E. Zinc absorption from composite meals. I. The significance of whest extraction rate, zinc, calcium, and protein content in meals based on bread. *Am. J. Clin. Nutr.* 1980, 33, 739–745. [CrossRef]
- Sandström, B.; Sandberg, A.S. Inhibitory effects of isolated inositol phosphates on zinc absorption in humans. J. Trace. Elem. Electrolytes Health Dis. 1992, 6, 99–103.
- Sandström, B.; Cederblad, A. Zinc absorption from composite meals. II. Influence of the main protein source. *Am. J. Clin. Nutr.* 1980, 33, 1778–1783. [CrossRef] [PubMed]
- Sandström, B.; Almgren, A.; Kivistö, B.; Cederblad, A. Effect of protein level and protein source on zinc absorption in humans. J. Nutr. 1989, 119, 48–53. [CrossRef]
- 42. Wood, R.J. Assessment of marginal zinc status in humans. J. Nutr. 2000, 130, 1350s–1354s. [CrossRef]
- Hotz, C.; Peerson, J.M.; Brown, K.H. Suggested lower cutoffs of serum zinc concentrations for assessing zinc status: Reanalysis of the second National Health and Nutrition Examination Survey data (1976–1980). Am. J. Clin. Nutr. 2003, 78, 756–764. [CrossRef]
- 44. Karr, M.; Mira, M.; Causer, J.; Earl, J.; Alperstein, G.; Wood, F.; Fett, M.J.; Coakley, J. Age-specific reference intervals for plasma vitamins A, E and beta-carotene and for serum zinc, retinol-binding protein and prealbumin for Sydney children aged 9–62 months. *Int. J. Vitam. Nutr. Res.* **1997**, *67*, 432–436.
- 45. Van Biervliet, S.; Van Biervliet, J.P.; Bernard, D.; Vercaemst, R.; Blaton, V. Serum zinc in healthy Belgian children. *Biol. Trace Elem. Res.* **2003**, *94*, 33–40. [CrossRef]
- Hess, S.Y.; Peerson, J.M.; King, J.C.; Brown, K.H. Use of serum zinc concentration as an indicator of population zinc status. *Food Nutr. Bull.* 2007, 28, S403–S429. [CrossRef] [PubMed]
- Wessells, K.R.; King, J.C.; Brown, K.H. Development of a plasma zinc concentration cutoff to identify individuals with severe zinc deficiency based on results from adults undergoing experimental severe dietary zinc restriction and individuals with acrodermatitis enteropathica. J. Nutr. 2014, 144, 1204–1210. [CrossRef] [PubMed]
- 48. Lowe, N.M.; Fekete, K.; Decsi, T. Methods of assessment of zinc status in humans: A systematic review. *Am. J. Clin. Nutr.* 2009, *89*, 2040s–2051s. [CrossRef] [PubMed]